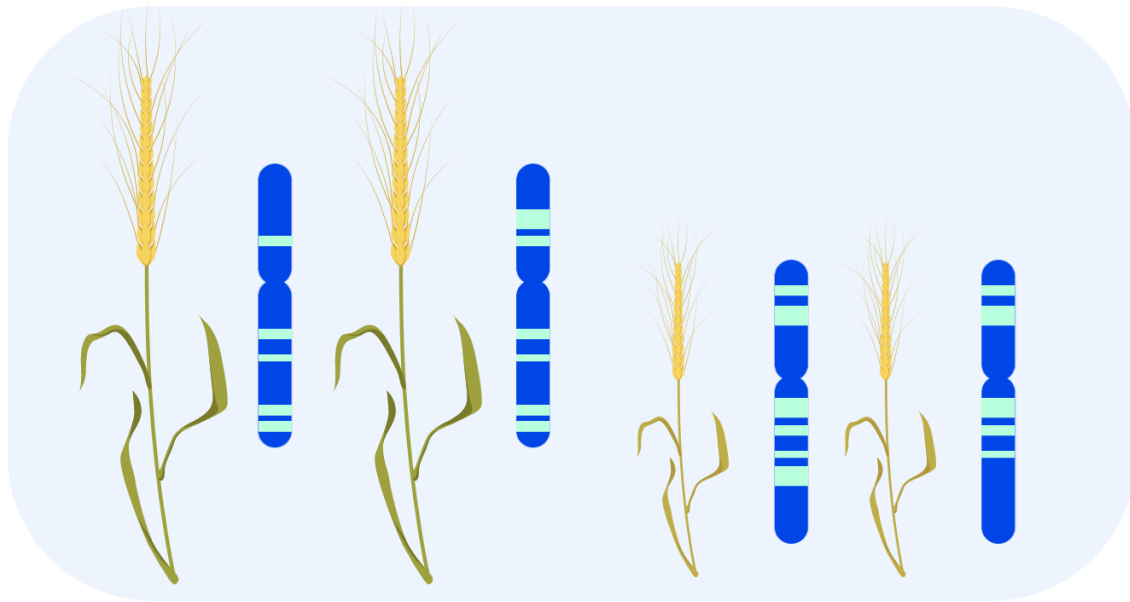


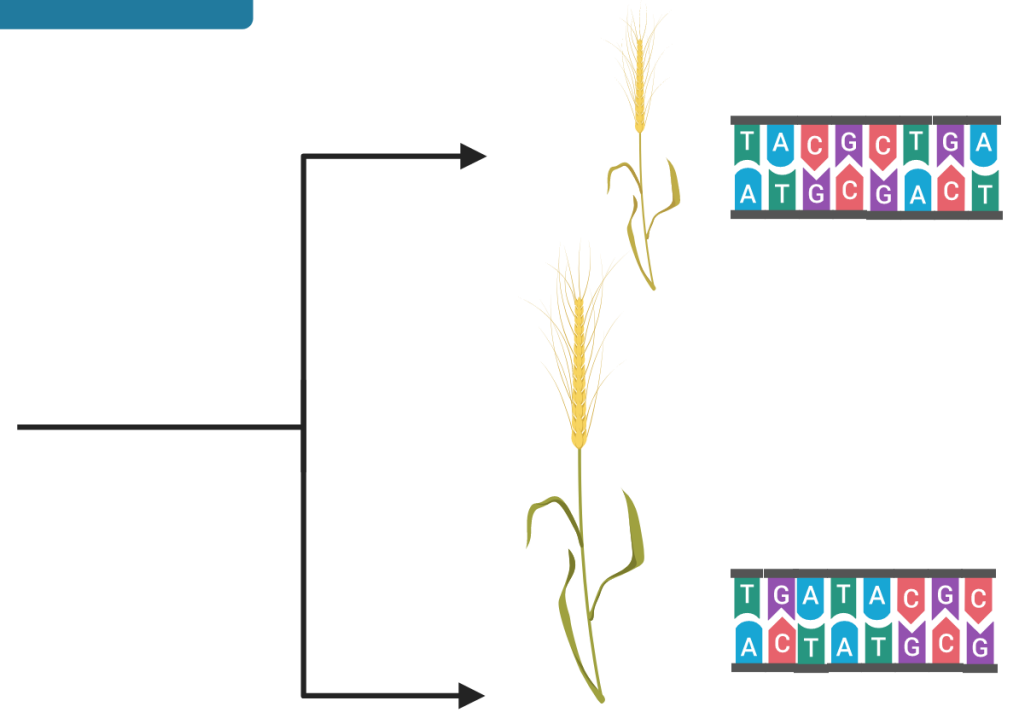
# Fine mapping and cloning genes

Ellie Taagen & Shantel A Martinez

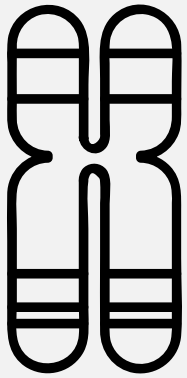
October 24, 2019



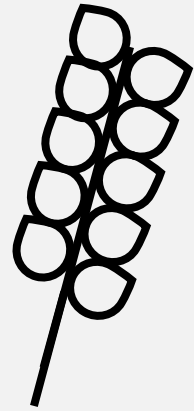
From phenotype and genotype associations



To identification of a trait's causal variant sequence



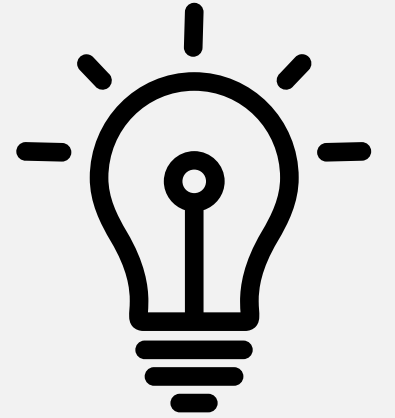
Background  
and Basics



Practical  
Example



Conceptual  
Activity

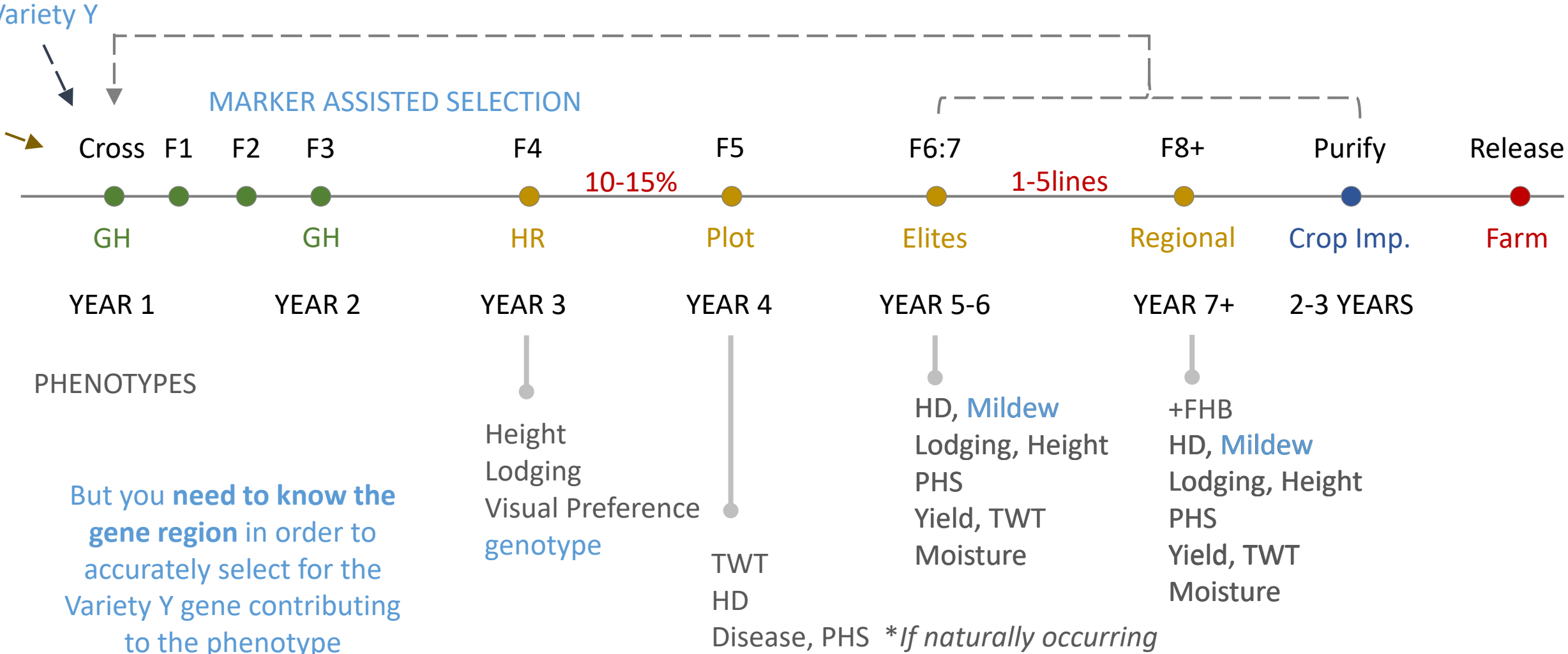


Challenges  
and  
Solutions

Fine mapping and cloning is a method for identifying causal genetic variants driving phenotypic variants

This allows for precise selection for desirable traits in the breeding germplasm

# Application: An example of traditional wheat breeding in Northeast



# Definitions of **key** concepts

## Genetic linkage analysis

Use statistical probability to test whether a phenotype is associated with a genetic marker by chance or because the phenotype's causal variant and the genetic marker are closely located in the same genomic segment

- The phenotype is something you can measure: disease susceptibility, grain weight, gene expression, etc.
- The resolution of the linkage analysis is limited by the distribution of the genetic markers and position of genetic recombination events

## Positional cloning

Based on the *physical position* of genetic sequence on chromosome, identify the gene and characterize the causal variant base pair(s) responsible for the phenotype

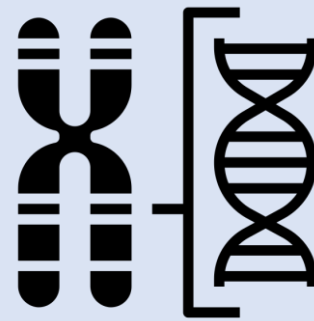
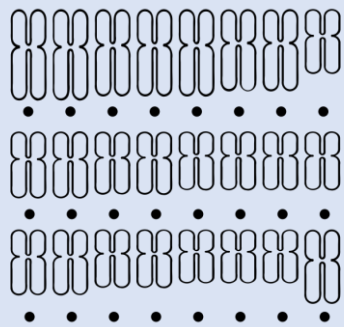
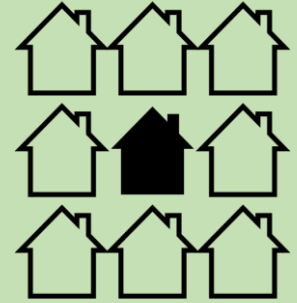
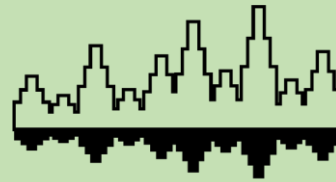
- Create a perfect marker for screening breeder's germplasm
- Vector based cloning system
  - DNA targeting tools such as the CRISPR-Cas9 system can facilitate more rapid introgression

*Correlation*

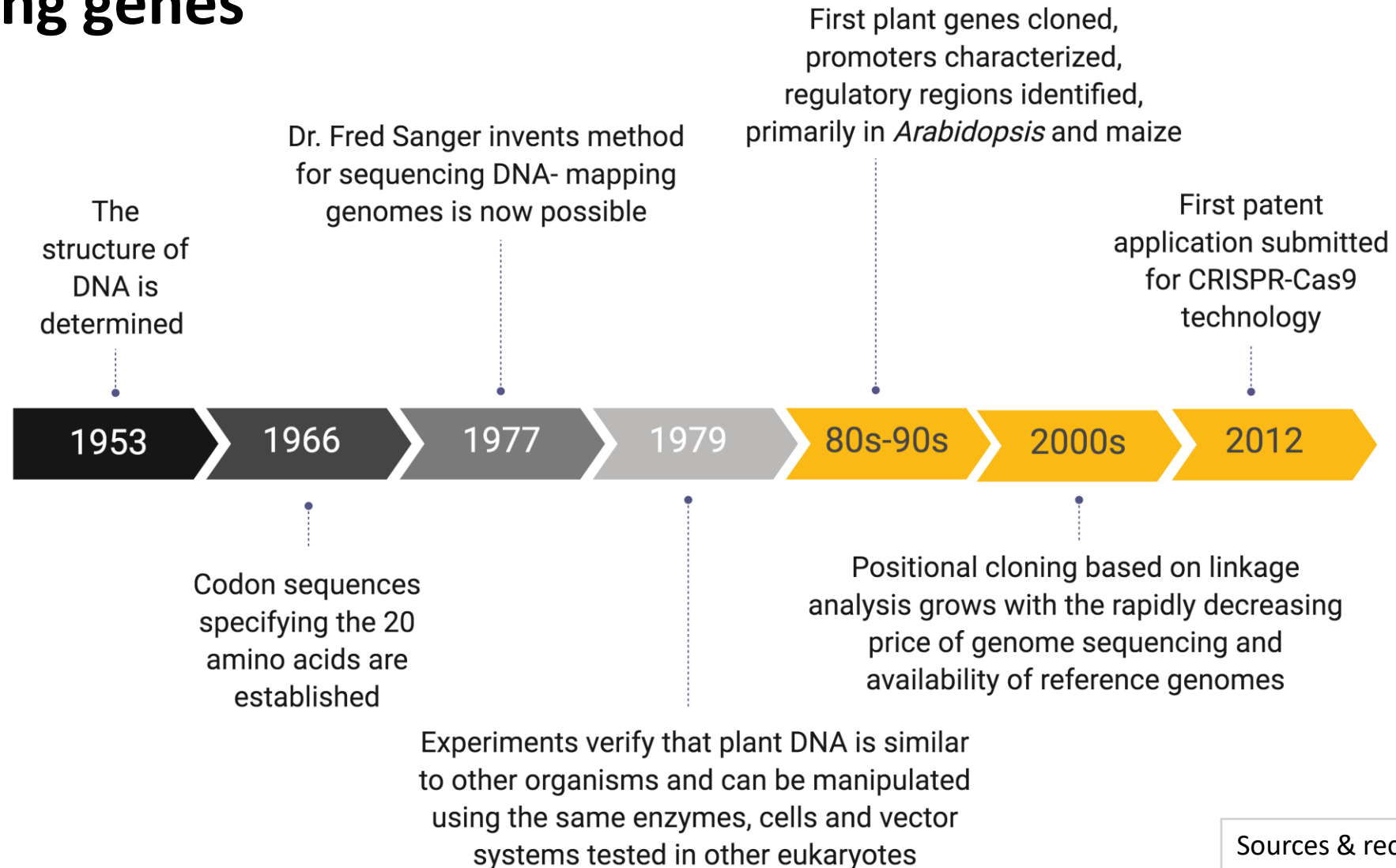
*Causation*



# The logic of mapping a gene



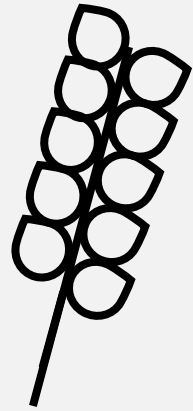
# A brief history of mapping and cloning genes



Sources & recommended reading  
*Goldberg, Plant Physiology, 2001*  
*The Gene, Siddhartha Mukherjee*



Background  
and Basics



Practical  
Example

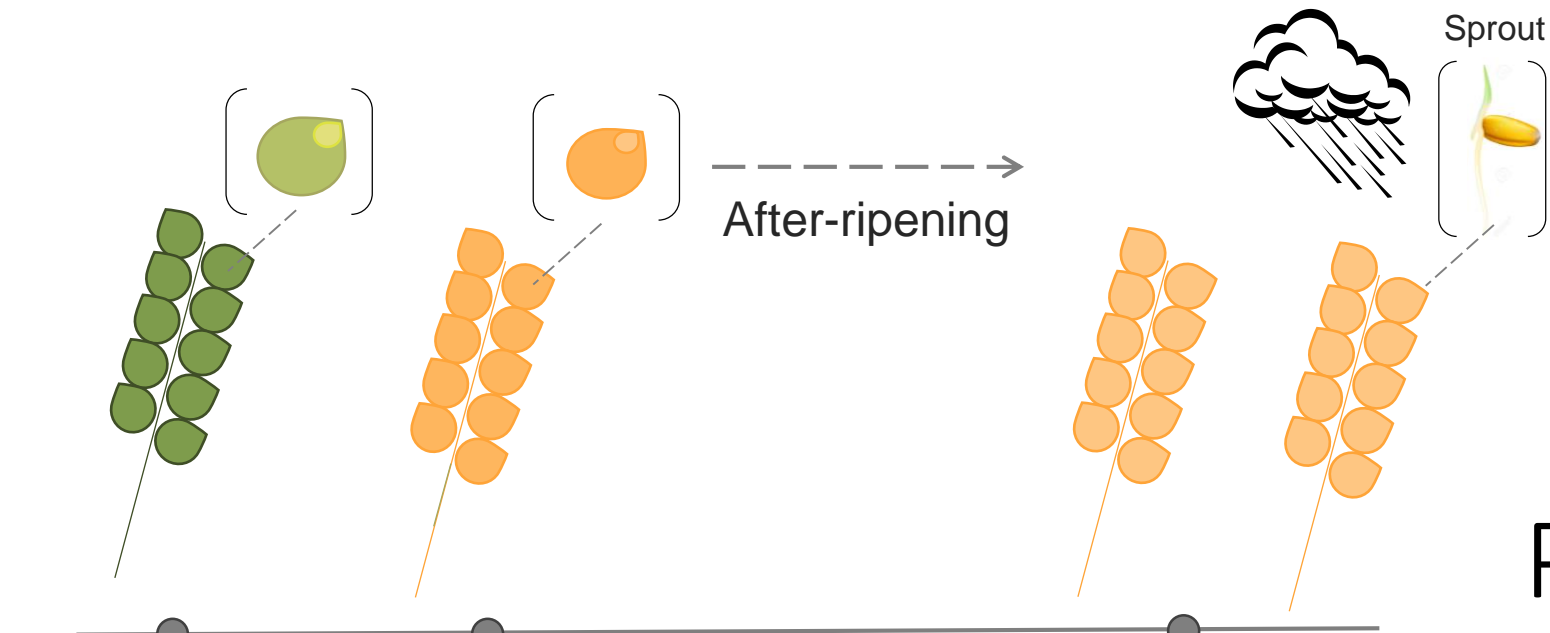


Conceptual  
Activity



Challenges  
and  
Solutions





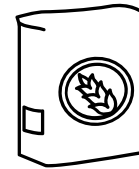
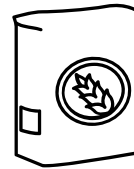
Physiological Maturity (PM)



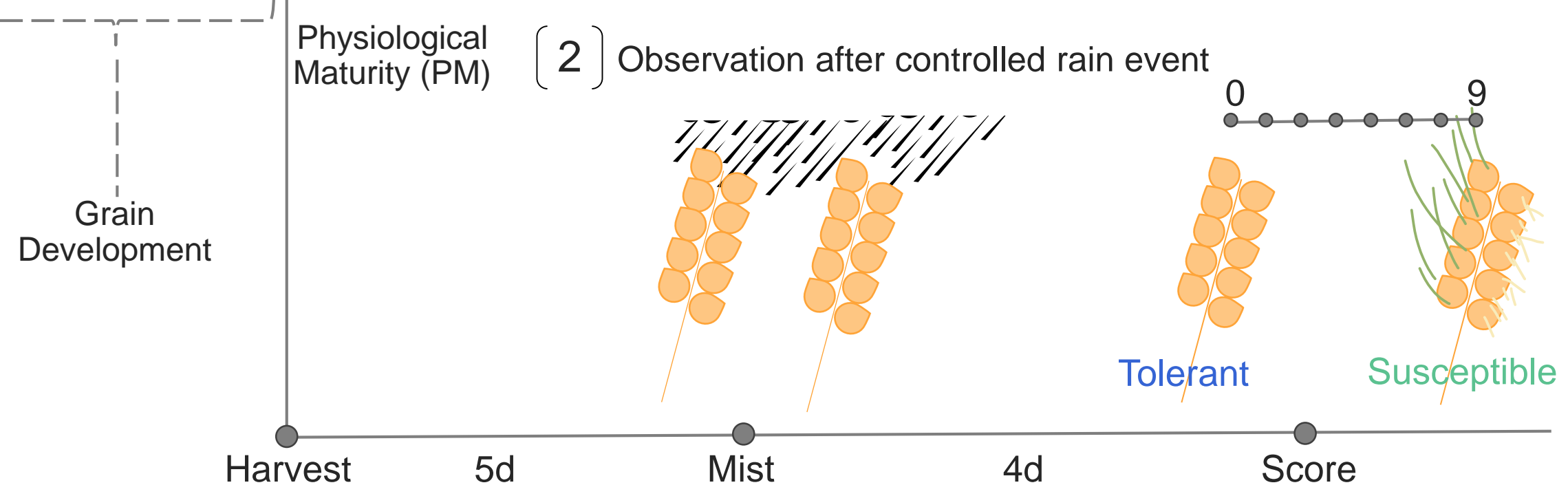
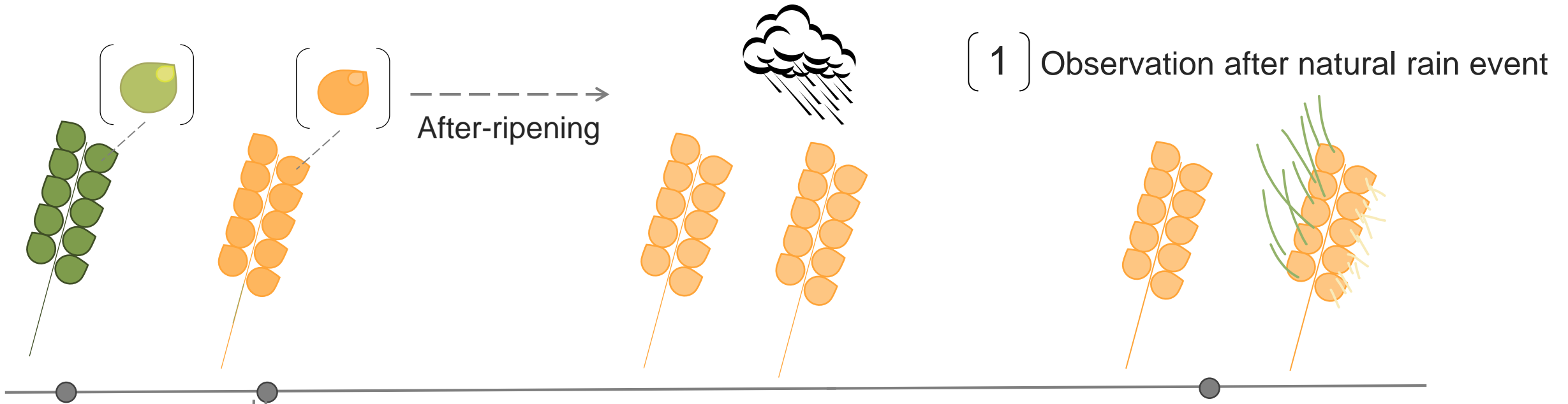
Combine Harvest

# Preharvest Sprouting

Grain Development



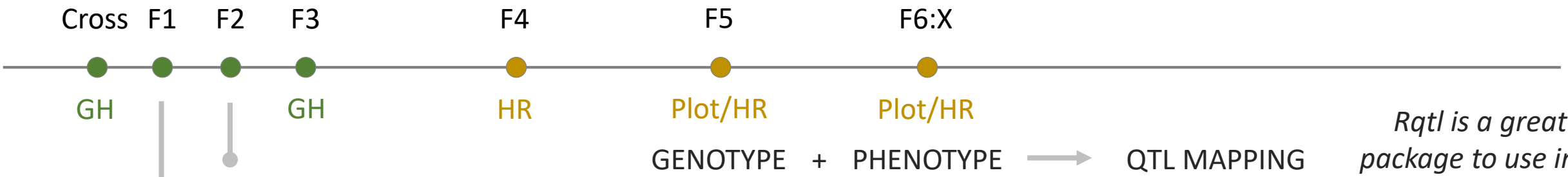
Decrease in Quality



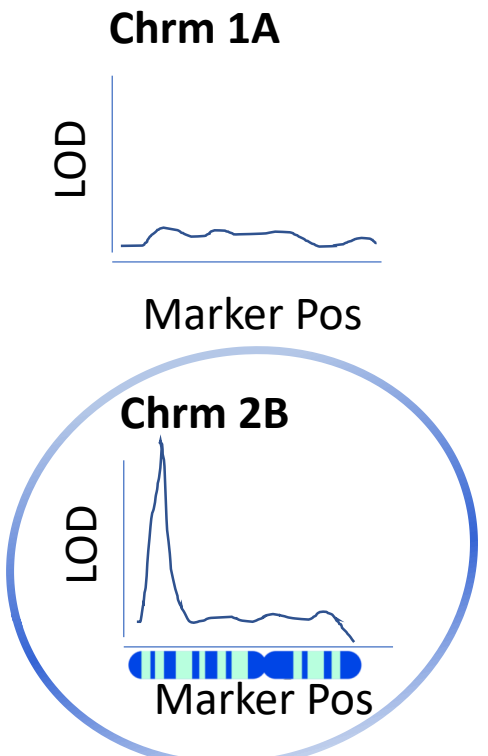
# QTL mapping a PHS tolerance gene

Identifies statistical association of the phenotype with the genotype

Tolerant Line Y X Susceptible Line Z

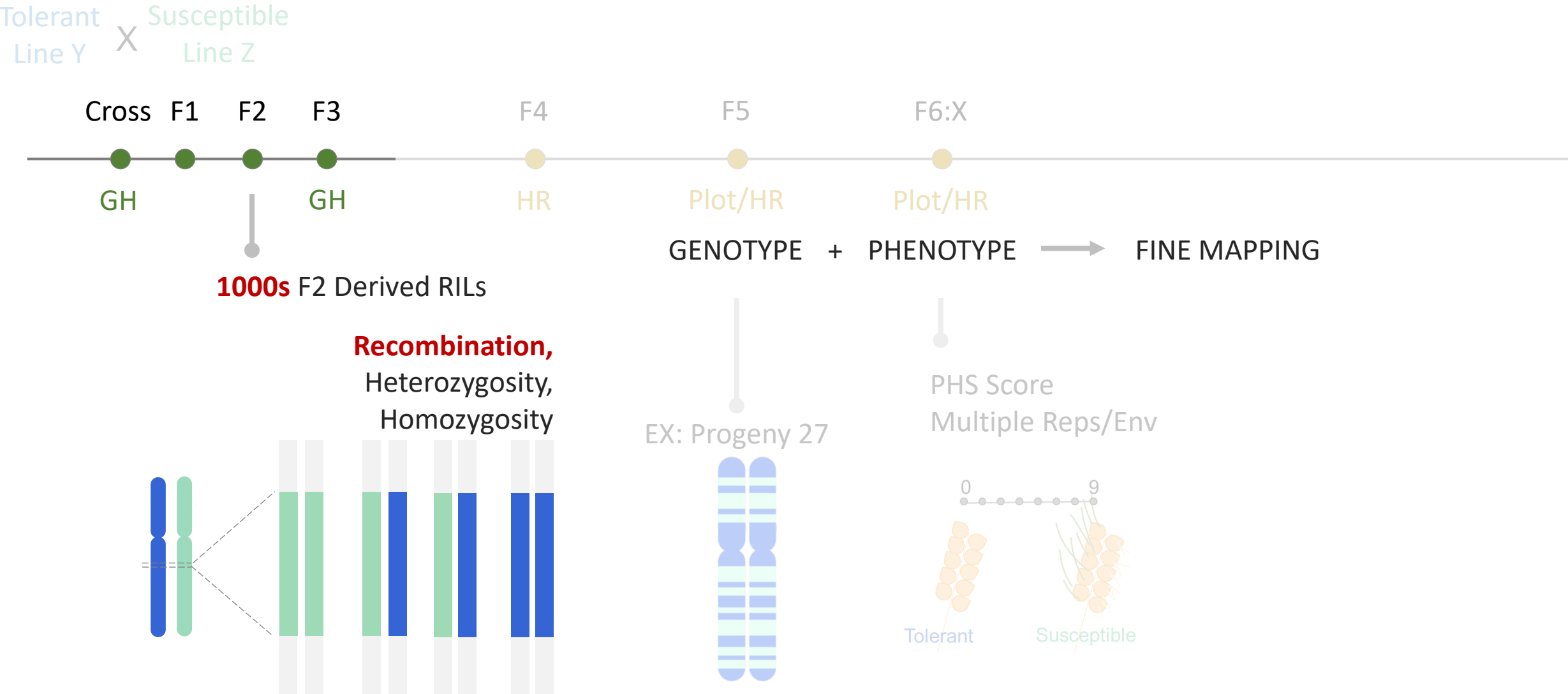


*Rqtl is a great package to use in R*



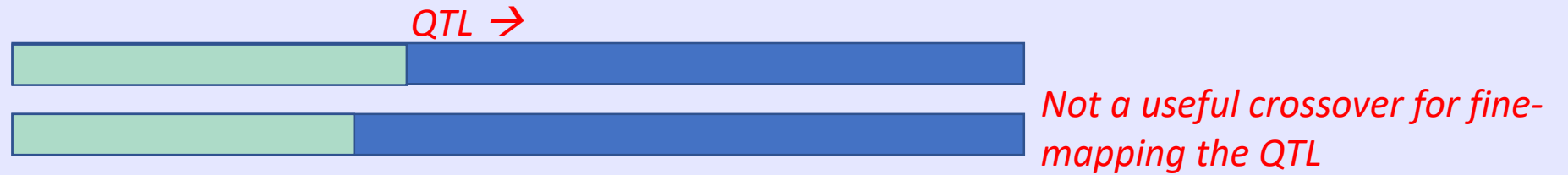


# Fine mapping the 2B QTL



## Why does the mapping population **size** matter?

- There are only 1-3 crossovers per chromosome per meiosis
  - The chance that a crossover happens somewhere informative for fine mapping the QTL decreases as we zoom-in!
  - A larger population can better measure the variability of the phenotype



## Why does the mapping population **structure** matter?

- Biparental (generally)

Select for heterozygosity →

CO provide finer resolution of the QTL →

heterogeneous inbred families (HIFs)



Background  
and Basics



Practical  
Example



Conceptual  
Activity



Challenges  
and  
Solutions

# Step 1: which varieties are PHS tolerant/susceptible?

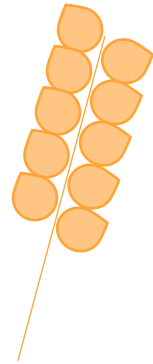
PHS Tolerant ← → PHS Susceptible



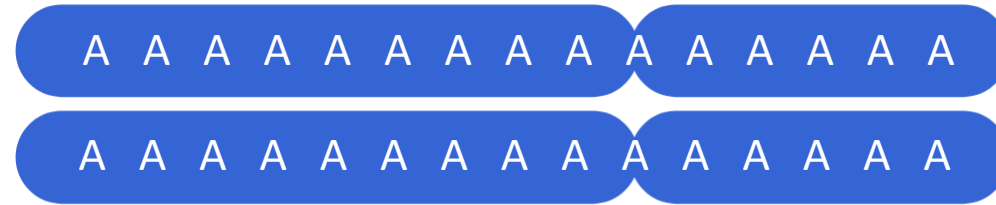


# Step 1: which varieties are PHS tolerant/susceptible?

Tolerant

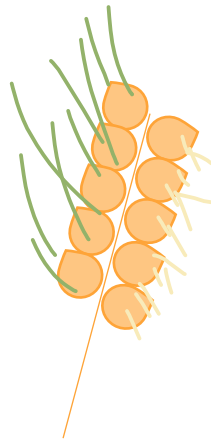


2B

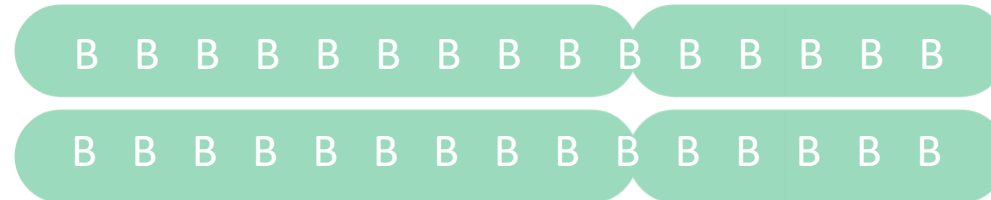


Causal Variant

Susceptible

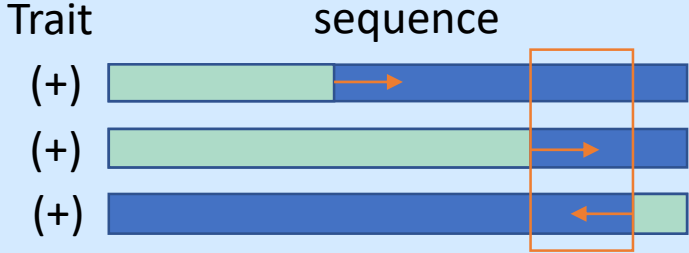
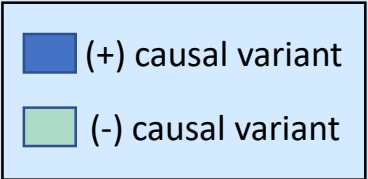
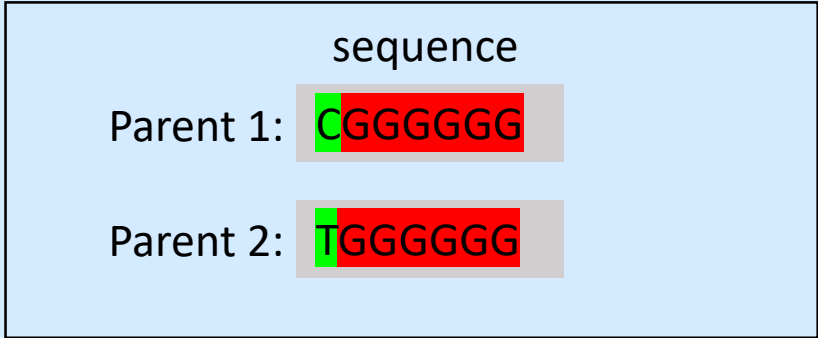
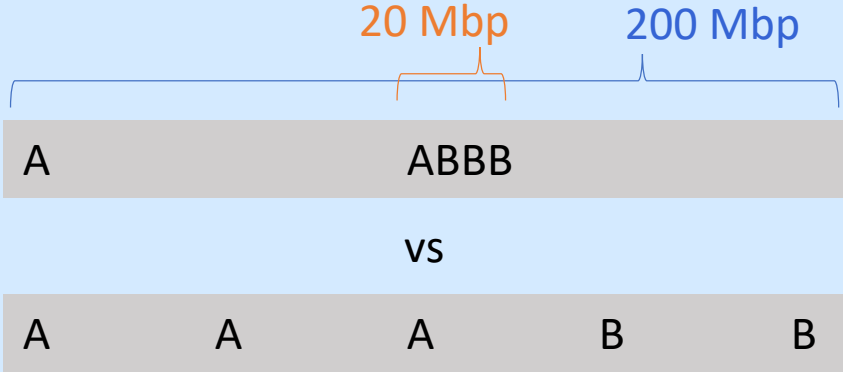


2B



# Genetic resolution can be a limiting factor


- Distribution of genetic markers (ie. SNP)
- Polymorphism
- Crossovers



## Step 2: order the lines into groups based on genotypes

100 Mbp	200 Mbp	300 Mbp	400 Mbp	500 Mbp	600 Mbp	700 Mbp
SNP	SNP	SNP	SNP	SNP	SNP	SNP
B	B	B	B	A	A	A

Recombination event  
(crossover event)

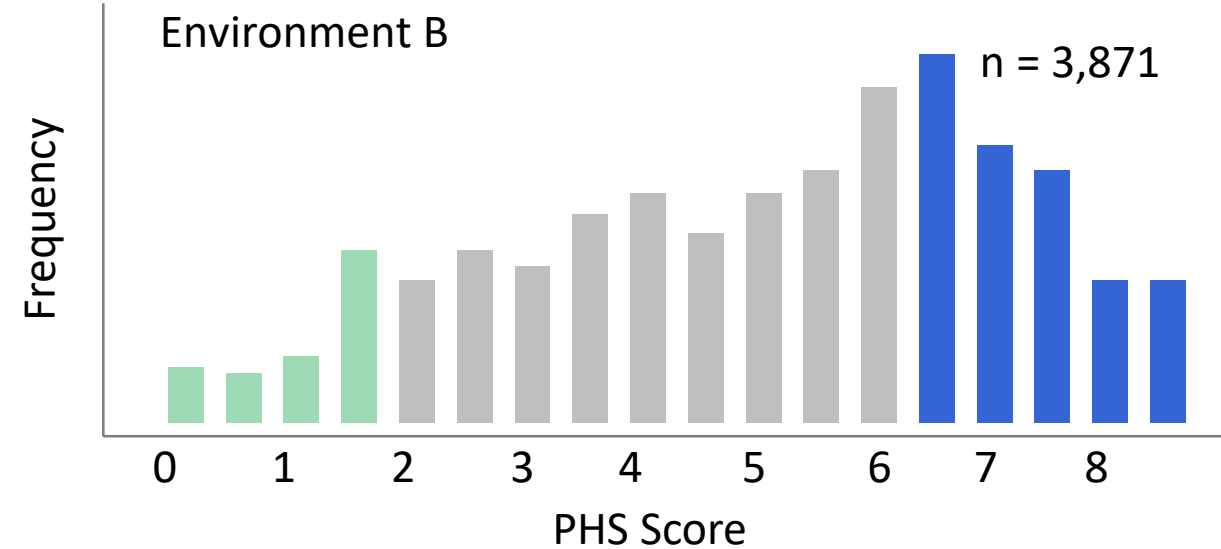
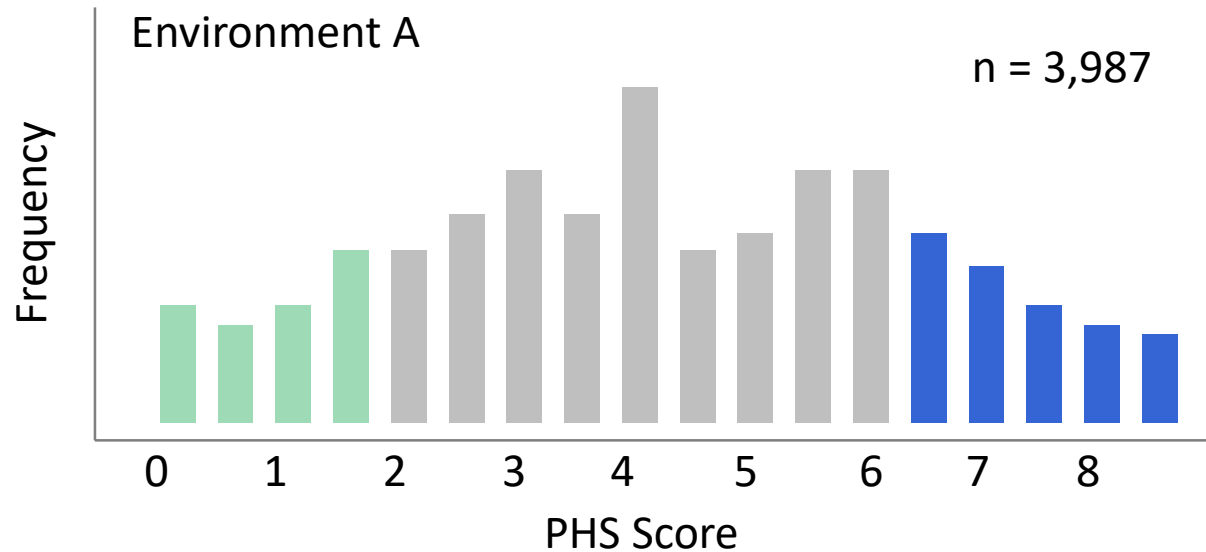


Do lines have similar recombination events?

Can the recombination events be ordered along the chromosome?

We suspect AAA lines to be tolerant and the BBB lines to susceptible but we don't know yet the about the recombinants

# Step 3: combine geno/pheno data



The handout is only looking at a small subset of these 4,000 progeny

Write the phenotype per line on the ordered genotypes strips



## Step 4: examine the progeny's geno/pheno

- What phenotype pattern do you see in association with the genotypes? Which haplotype is always susceptible? Tolerant?
- Can you identify where there is a critical crossover event? Highlight the two flanking genetic markers.

## Recombination event

Phenotype	100 Mbp SNP	200 Mbp SNP	300 Mbp SNP	400 Mbp SNP	500 Mbp SNP	600 Mbp SNP	700 Mbp SNP
6	B	B	B	B	A	A	A
1	A	A	A	A	A	A	B
2	A	A	A	A	A	A	B
1	A	A	A	B	B	B	B
2	A	A	A	B	B	B	B
8	B	A	A	A	A	A	A
2	B	A	A	A	A	A	A
2	A	B	B	B	B	B	B
1	A	B	B	B	B	B	B
7	A	B	B	B	B	B	B

Having Cayuga markers (A) **up until 700Mb** results in a Cayuga PHS Tolerant phenotype (1,2)

Same effect: **up to 400Mb**

Recombination event **between 100-200Mb** shows either a Tol or Susc phenotype

Suggesting that **a gene between 100-200Mb** is associated with PHS tolerance

?

However, we currently do not have enough markers in that region to see which gene in the region provides tolerance

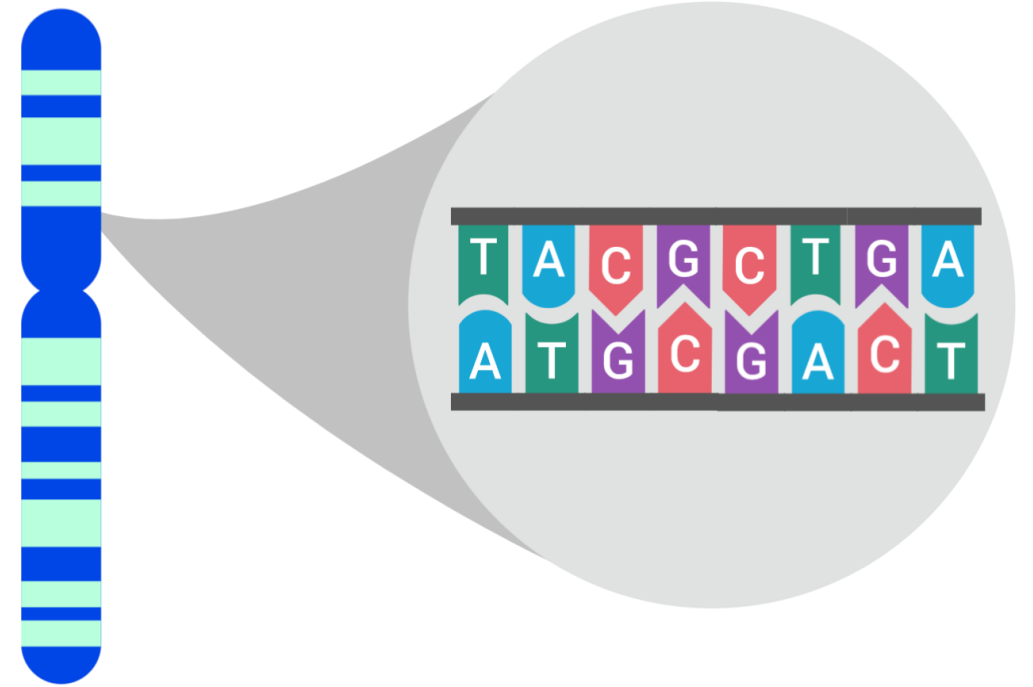
## Step 4: examine the progeny's geno/pheno

- What phenotype pattern do you see in association with the genotypes? Which haplotype is always susceptible? Tolerant?
- Can you identify where there is a critical crossover event? Highlight the two flanking genetic markers.
- You don't have the resources to phenotype everything - so pick only the lines that have a crossover between to critical flanks or are heterozygous



# Designing genetic markers

We want to zoom in on the sequence between the critical flanking positions ...  
...AKA identify positions where we can design genetic markers!



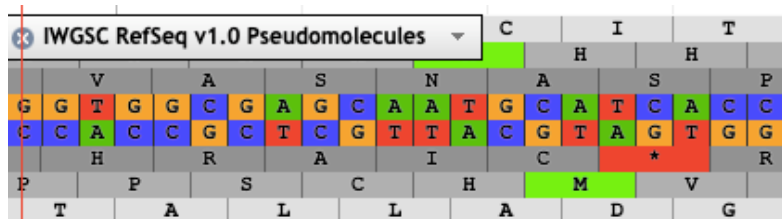
Useful genetic markers are both:

- Polymorphic for your parent lines
- Chromosome specific (and genome specific for polyploid species)

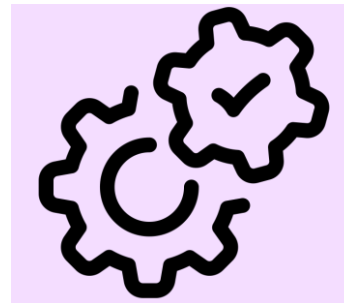
# Designing genetic markers

- We put our resources into developing 10 new genetic markers between the critical flanking positions

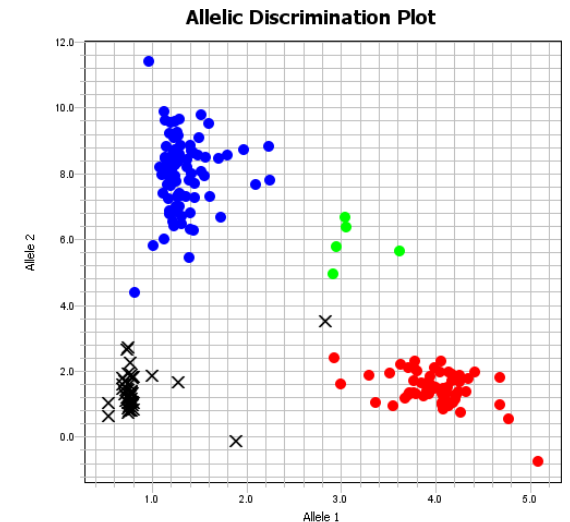
*Quick how-to for KASP markers:*



After you ID a polymorphic & site specific SNP, ie. 90K, grab sequence surrounding SNP from reference genome



Use tools like [Oligo-Calc](#) and [NIH Primer Blast](#) to optimize primer design



Test the markers- 3 primer fluorescent system detects alleles A, B and AB

# Step 5: the mapping process is iterative

- We genotype the progeny of the plants we advanced with the 10 new markers and measure their PHS phenotype
- Look at your new results - what phenotype pattern do you see in association with the genotypes? Which haplotype is always susceptible? Tolerant?
- Can you identify where there is a critical crossover event? Highlight the two flanking genetic markers.

# Step 6: prioritize list of genes

- A: Look at the **polymorphism between your parental lines** in the [EcoTILLING](#) browser.  
Prioritize genes that have truncation mutations or changes with negative BLOSUM 62 scores in conserved regions (THIS CAN MISS REGULATORY DIFFERENCES)
- B: Look within **expression databases** for the expression profile of the genes.  
[expVIP](#) [WheatExp](#) [Wheat eFP](#)  
Prioritize genes that are expressed in your target tissue and developmental time
- C: Find the **collinear** and closest genes in Rice, Brachy, Barley, Maize.  
Find the closest gene in Arabidopsis. Record the names and numbers. If there are close paralogs, you may need to generate a phylogenetic tree to find the closest homologs.  
Do a literature search to learn about the function of this gene or gene family:  
Starting with barley->brachy->rice->maize and then Arabidopsis.  
Transcription factors are genes regulating protein stability are always good candidates  
Most of the QTLs cloned in wheat so far are in these categories
- D: Prepare a small **document** for each gene describing:  
Conserved domains, function, references, expression profile,  
polymorphisms between parental lines.  
Integrate that info into a Table and prioritize your list

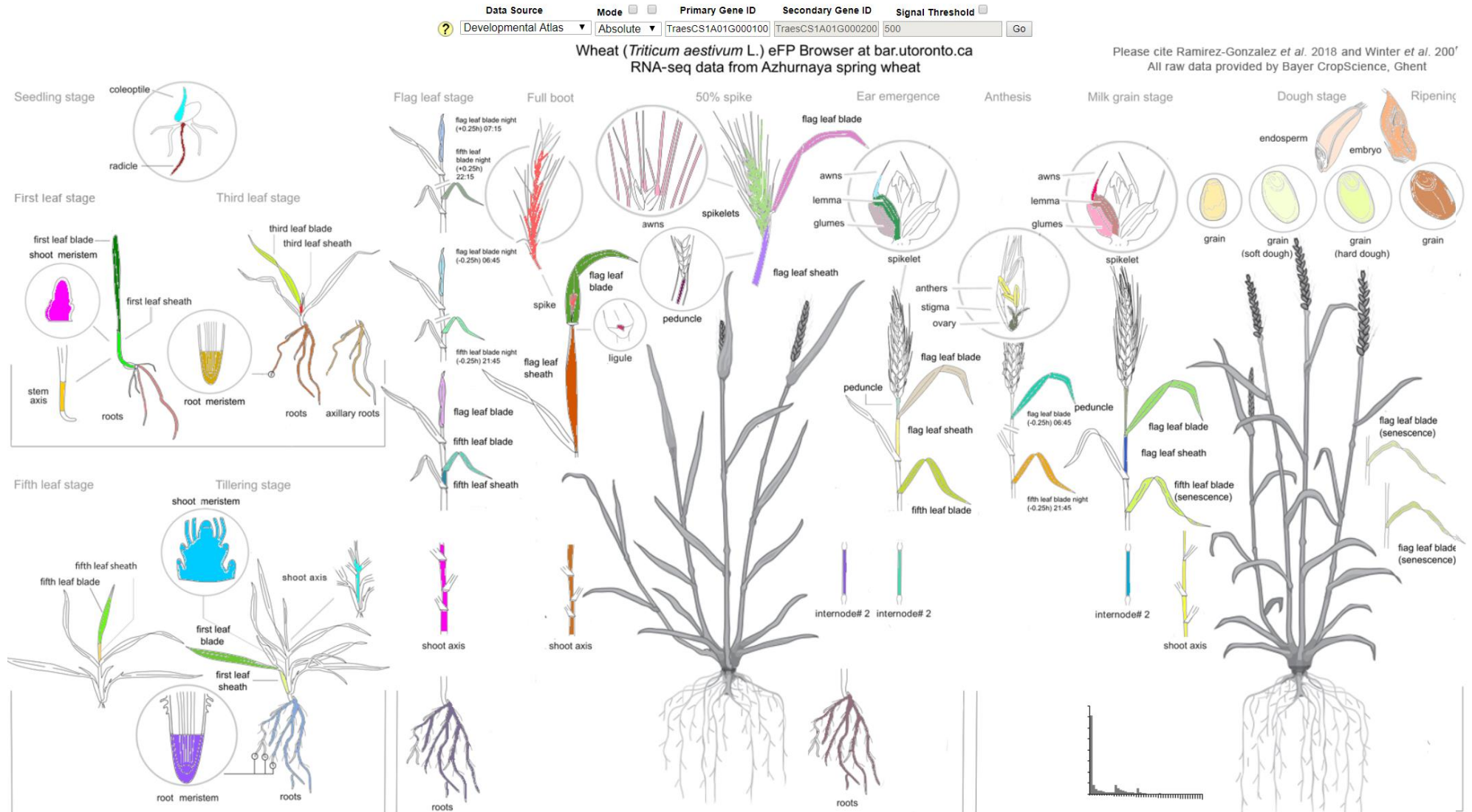
# Step 6: prioritize list of genes

[bit.ly/35UqEXP](http://bit.ly/35UqEXP)

Search the gene name(s):

Do some of the genes show no expression (yellow) in the tissue you would expect it to be present in?

Check the entire list of genes

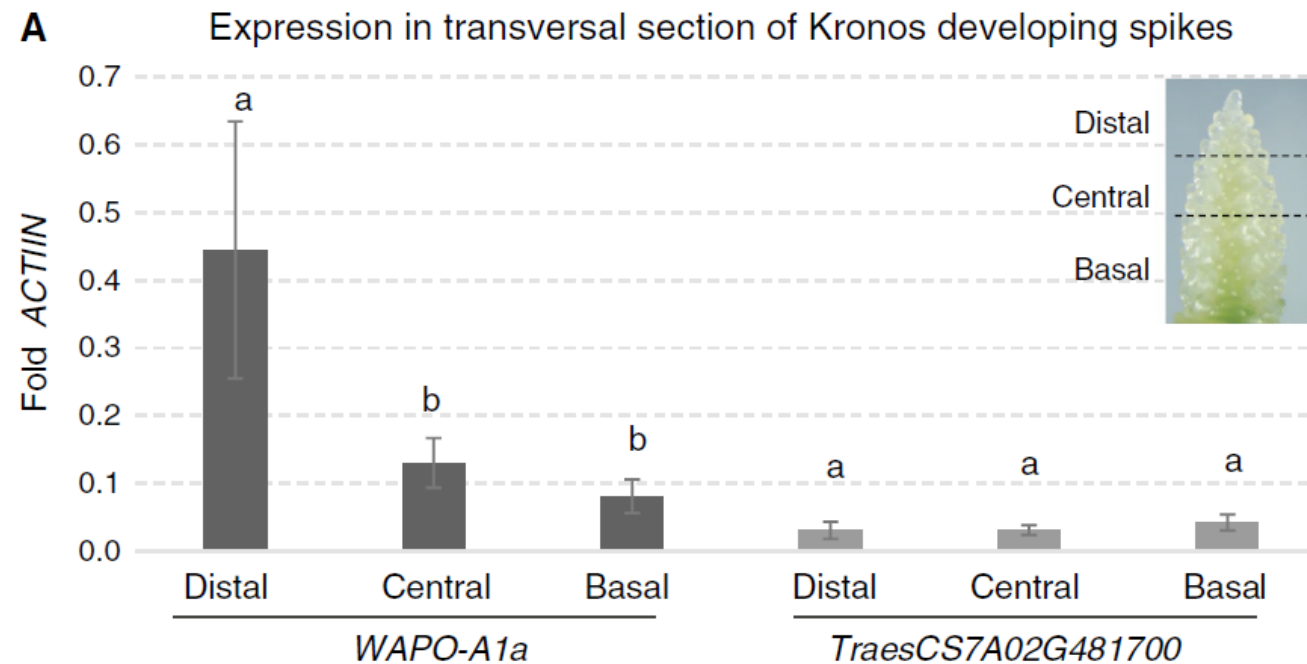


[HANDOUT 5: GENE LIST]

# Step 7: gene validation

Check for gene expression within your lines and tissue relevant to the trait

**Fig. 4** Transcript levels in developing spikes at the floret primordia stage relative to *ACTIN* as endogenous control. **a** *WAPO-A1* and *TraesCS7A02G481700* in basal, central and distal sections of Kronos developing spikes. The experiment was repeated twice, and data were analyzed together using experiment as block. Means within each gene were compared using Tukey tests ( $=0.05$ ). Bars are SE of the means. **b** Transcript levels of *WAPO-A1* in hexaploid near isogenic HIF lines homozygous for the *WAPO-A1b* and *WAPO-*



# Step 7: gene validation

- You look at your research budget and there is only enough funding to pursue **three** genes for knockouts with CRISPR-Cas9



- Pick the plants you want to transform: which allele are we trying to knockout?
- What phenotype do you expect?

# Step 7: gene validation

Gene A

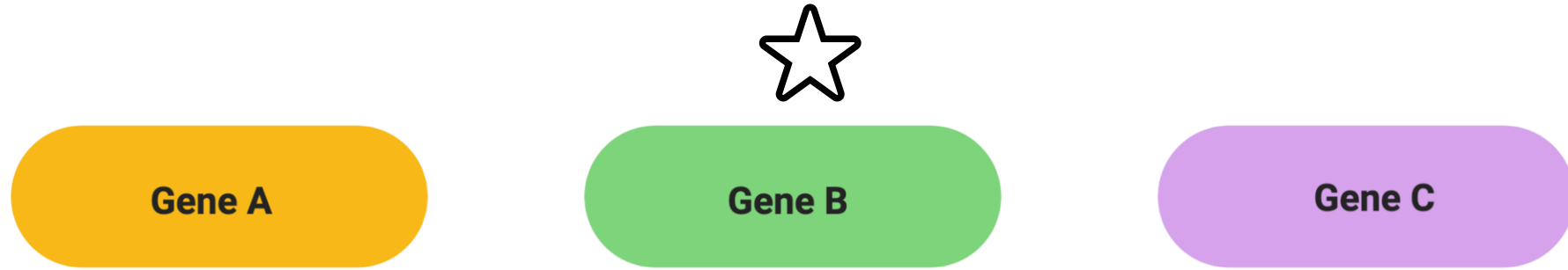
Gene B

Gene C

Phenotype	Treatment	Treatment	Treatment



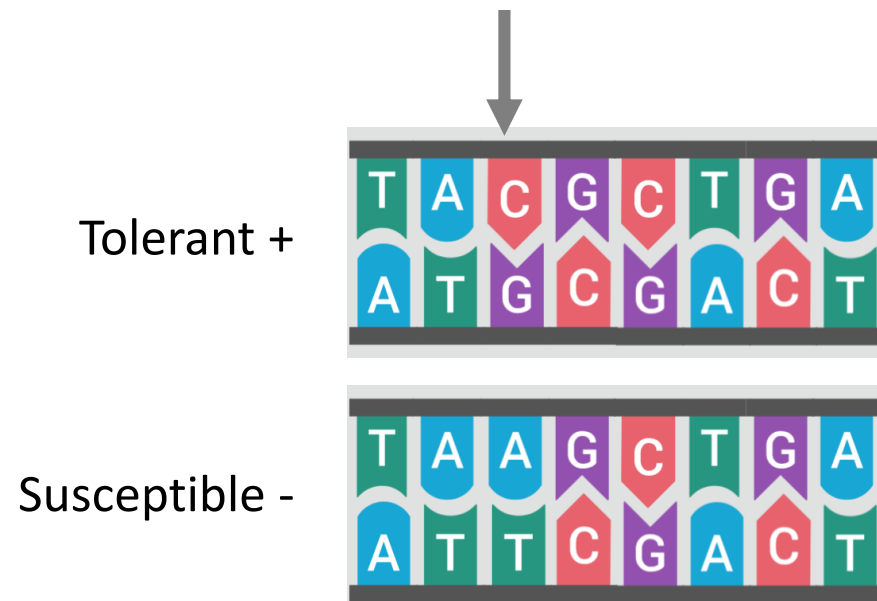
# Step 7: gene validation



Phenotype	Treatment	Treatment	Treatment
1	KO	WT	WT
7	WT	KO	WT
2	WT	WT	KO

# Step 8: positional cloning

- Sequence the gene from both + and - phenotype
- Identify the causal variant in the sequence (ie. is it a nucleotide shift in the promoter)



```

TaSdr-B1a GCCCTTCACTGGACGCCACTGGAATCCACAGTCTCTCCCTCCAAAGCAGCGCGCCCCG 60
TaSdr-B1b GCCCTTCACTGGACGCCACTGGAATCCACAGTCTCTCCCTCCAAAGCAGCGCGCCCCG 60
                Sdr-5 Forward primer
TaSdr-B1a GACTCGCCTCCGCTACGTGTCGGCCCCGTCCCGCCCGCTCGCCCACGTACCCCGCGCC 120
TaSdr-B1b GACTCGCCTCCGCCTACGTGTCGGCCCCGTCGCCCGCCCGCTCGCCCACGTACCCCGCGCC 120
                PmlI recognition site
TaSdr-B1a TCGTTCCCACGTGCCCTCCCTCTGCGGCATCCGATTGGCCGCCACGCCTTCTTAAG 180
TaSdr-B1b TCGTTCCCACGTGCCCTCCCTCTGCGGCATCCGATTGGCCGCCACGCCTTCTTAAG 180

TaSdr-B1a CGGCACGGCACCGGGACCC AACGCCGTGCACTCCGTCCACCCCGTCAGCAGACTTCGAC 240
TaSdr-B1b CGGCACGGCACCGGGACCC AACGCCGTGCACTCCGTCCACCCCGTCAGCAGACTTCGAC 240
                PmlI recognition site  initiation codon
TaSdr-B1a TCGCACGTGCACGCAATGGCCATGGTGCAGCCGGCGGACATGGCCGTC AAGGCCAACGAG 300
TaSdr-B1b TCGCCGTGCCACGCAATGGCCATGGTGCAGCCGGCGGACATGGCCGTC AAGGCCAACGAG 300

TaSdr-B1a ATCTCGCGGGTTCGGGCCATCGCGCCAAAGCCCGCCTGCCGCGCTCGCCGGTGCAG 360
TaSdr-B1b ATCTCGCGGGTTCGGGCCATCGCGCCAAAGCCCGCCTGCCGCGCTCGCCGGTGCAG 360

TaSdr-B1a GCGATCGACGGCGCCCGGACCCGCGTGTCTGCCACCTGCAGAACAGGCCGTGCCGGGA 420
TaSdr-B1b GCGATCGACGGCGCCCGGACCCGCGTGTCTGCCACCTGCAGAACAGGCCGTGCCGGGA 420

TaSdr-B1a AGGAAGCGGGGGCGCCGAGCGCCGTGCCGGTGTCCGCGCCGCCGCTGCCGCCAAGAGG 480
TaSdr-B1b AGGAAGCGGGGGCGCCGAGCGCCGTGCCGGTGTCCGCGCCGCCGCTGCCGCCAAGAGG 480

TaSdr-B1a AAGAGGGCGGGTACCCGGTGCCTCCGGTGCAGGGGGCGGGCCACCGACGCGGTG 540
TaSdr-B1b AAGAGGGCGGGTACCCGGTGCCTCCGGTGCAGGGGGCGGGCCACCGACGCGGTG 540

TaSdr-B1a GTGTCCACCGCGACGAGGGCTATGTGTCCTGCCGGGAGTGCATGCATGCCGTTTGGC 600
TaSdr-B1b GTGTCCACCGCGACGAGGGCTATGTGTCCTGCCGGGAGTGCATGCATGCCGTTTGGC 600

TaSdr-B1a TCCCTCCC GCCGCGACCCGCGAGTACCCGGGGAATCTGACGATGCTCTCGACGACCATG 660
TaSdr-B1b TCCCTCCC GCCGCGACCCGCGAGTACCCGGGGAATCTGACGATGCTCTCGACGACCATG 660

TaSdr-B1a GTGGCGGGCAGCAGCAAGAGGAGGAGAGGGACGTCCCGTGGAGCGGACCTGTGCGG 720
TaSdr-B1b GTGGCGGGCAGCAGCAAGAGGAGGAGAGGGACGTCCCGTGGAGCGGACCTGTGCGG 720

TaSdr-B1a AAGTGTGGAGCCAAAGGTGATCTCGCCGCGGGGATGCGCCCGTAGGATCCACCATC 780
TaSdr-B1b AAGTGTGGAGCCAAAGGTGATCTCGCCGCGGGGATGCGCCCGTAGGATCCACCATC 780

TaSdr-B1a CACGTGCAATCCATCGTCCCGGCGCCGTCGACGCGACCAGCACCGCCCTCGAAGACG 840
TaSdr-B1b CACGTGCAATCCATCGTCCCGGCGCCGTCGACGCGACCAGCACCGCCCTCGAAGACG 840
                Sdr-5 Reverse primer
TaSdr-B1a GCGGAGGAGGTGGAGGGCGAGGTGGAGACCGACCGCTCCCGGGGTCGTCACGGACTCG 900
TaSdr-B1b GCGGAGGAGGTGGAGGGCGAGGTGGAGACCGACCGCTCCCGGGGTCGTCACGGACTCG 900

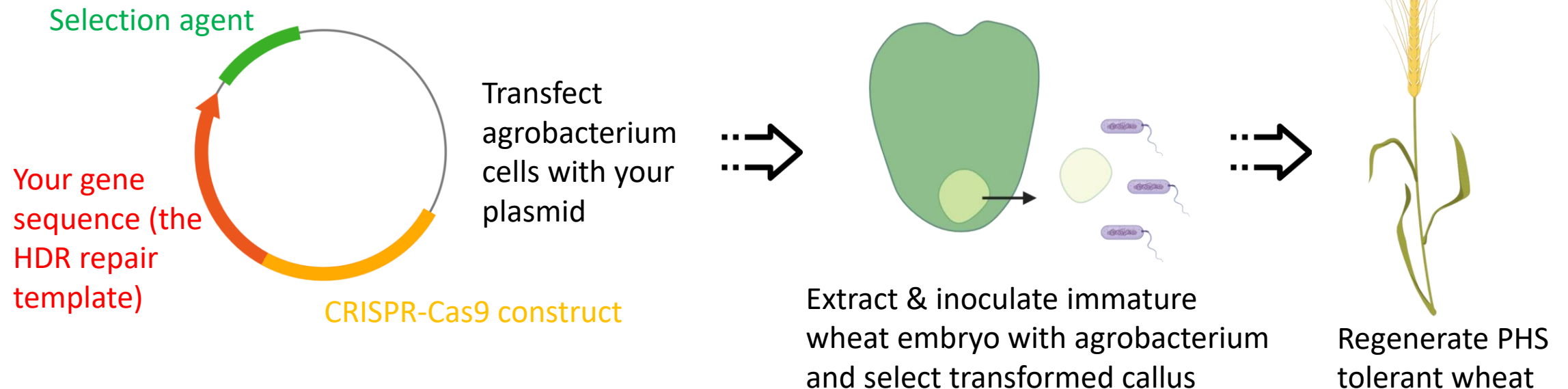
TaSdr-B1a AGCAACCGCTCCGGTGGTGAACGACGCGTACAAGGAGATGGTGGCGCGCCGAGTGC 960
TaSdr-B1b AGCAACCGCTCCGGTGGTGAACGACGCGTACAAGGAGATGGTGGCGCGCCGAGTGC 960

TaSdr-B1a CTGTGGCTCGGCGCGGTGGCCGCTCGAGGAGGATCAGCGGGAGGTGGCGCTGGTGGT 1020
TaSdr-B1b CTGTGGCTCGGCGCGGTGGCCGCTCGAGGAGGATCAGCGGGAGGTGGCGCTGGTGGT 1020

```

# Step 9: is my gene sufficient to determine the phenotype?

- Create a HDR construct with your tolerant gene sequence and use agrobacterium to knock-in the trait into a PHS susceptible variety





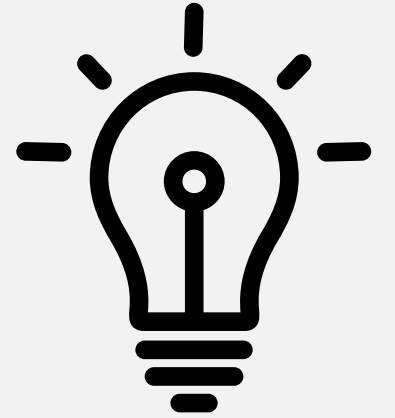
Background  
and Basics



Practical  
Example

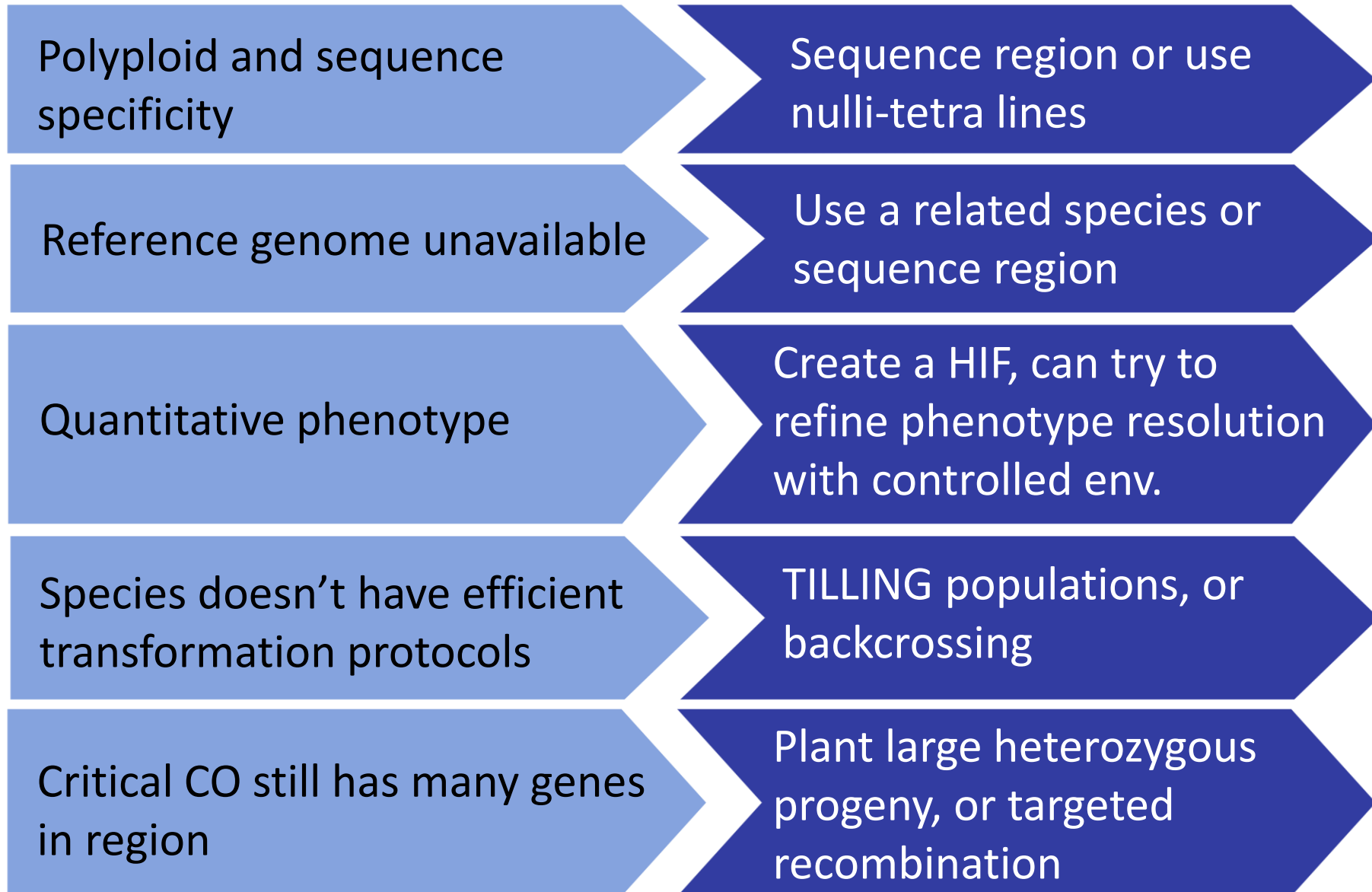


Conceptual  
Activity

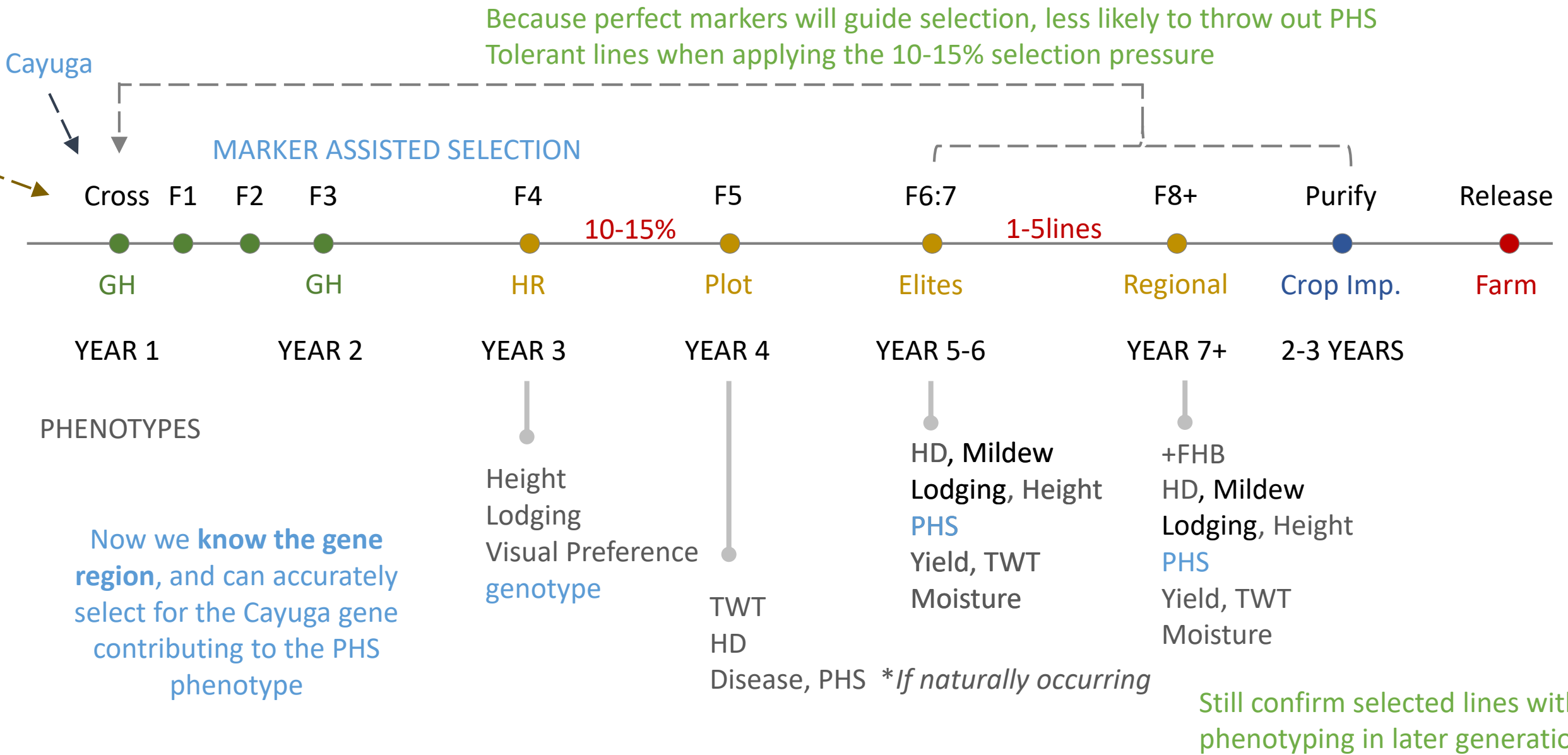


Challenges  
and  
Solutions

# Challenges / Solutions



# Breeding after fine mapping & cloning

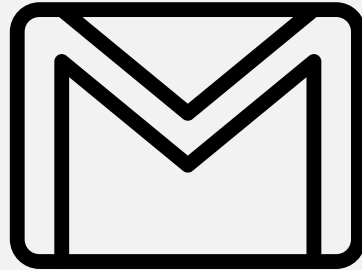


# LinkedIn



ella-taagen  
shantel-a-martinez

# Email



et395  
sam594

# Slides/Resources



[shantel-martinez.github.io/talks](https://shantel-martinez.github.io/talks)

# REFERENCES

- Goldberg RB (2001) From Cot Curves to Genomics. How Gene Cloning Established New Concepts in Plant Biology. *Plant Physiology* 125:4–8. <https://doi.org/10.1104/pp.125.1.4>
- Kuzay S, Xu Y, Zhang J, et al (2019) Identification of a candidate gene for a QTL for spikelet number per spike on wheat chromosome arm 7AL by high-resolution genetic mapping. *Theor Appl Genet* 132:2689–2705. <https://doi.org/10.1007/s00122-019-03382-5>
- McMaster GJ, Derera NF (1976) Methodology and sample preparation when screening for sprouting damage in cereals. *Cereal Research Communications* 4:251–254
- Mukherjee S (2016) *The gene: an intimate history*, First Scribner hardcover edition. Scribner, New York
- Ramírez-González RH, Borrill P, Lang D, et al (2018) The transcriptional landscape of polyploid wheat. *Science* 361:eaar6089. <https://doi.org/10.1126/science.aar6089>
- Zhang Y, Miao X, Xia X, He Z (2014) Cloning of seed dormancy genes (TaSdr) associated with tolerance to pre-harvest sprouting in common wheat and development of a functional marker. *Theor Appl Genet* 127:855–866. <https://doi.org/10.1007/s00122-014-2262-6>



# MAPPING/CLONING RESOURCES

**Wheat genome sequence browser:** URGI Blast [https://urgi.versailles.inra.fr/blast\\_iwgsc/blast.php](https://urgi.versailles.inra.fr/blast_iwgsc/blast.php)

URGI JBrowse

[https://urgi.versailles.inra.fr/jbrowseiwgsc/gmod\\_jbrowse/?data=myData%2FIWGSC\\_RefSeq\\_v1.0&loc=chr1A%3A59558565..534840209&tracks=HighConfidenceGenesv1.0&highlight=](https://urgi.versailles.inra.fr/jbrowseiwgsc/gmod_jbrowse/?data=myData%2FIWGSC_RefSeq_v1.0&loc=chr1A%3A59558565..534840209&tracks=HighConfidenceGenesv1.0&highlight=)

**Primer Design:** Oligo-Calc <http://biotools.nubic.northwestern.edu/>

NIH Primer Blast <https://www.ncbi.nlm.nih.gov/tools/primer-blast/> PolyMarker <http://polymarker.tgac.ac.uk>

**SNP functional changes:** EcoTILLING browser <https://dubcovskylab.ucdavis.edu/eco-tilling-blast>

**Expression databases:** expVIP <http://www.wheat-expression.com> WheatExp <https://wheat.pw.usda.gov/WheatExp/>

Wheat eFP [http://bar.utoronto.ca/efp\\_wheat/cgi-bin/efpWeb.cgi](http://bar.utoronto.ca/efp_wheat/cgi-bin/efpWeb.cgi)

Ensembl Plants <https://plants.ensembl.org/index.html>

**Gene literature review:** Google Scholar <https://scholar.google.com> PubMed <https://www.ncbi.nlm.nih.gov/pubmed/>

**Design guide RNA:** Cermak et al., 2017 The Plant Cell <http://www.plantcell.org/content/29/6/1196>

Voytas Lab Plant Genome Engineering [http://cfans-pmorrell.oit.umn.edu/CRISPR\\_Multiplex/](http://cfans-pmorrell.oit.umn.edu/CRISPR_Multiplex/)

Wheat CRISPR <https://crispr.bioinfo.nrc.ca/WheatCrispr/>

**Sequence Alignment:** ClustalOmega <https://www.ebi.ac.uk/Tools/msa/clustalo/>

# HANDOUT RESOURCES

Google Sheet with handouts: [bit.ly/2JdxFIN](https://bit.ly/2JdxFIN)

## PRINTING INSTRUCTIONS:

PartA: Print Handout 1 for n groups (there is enough room for multiple per page)

PartB: Print Handout 2+3 for n groups; Cut apart Handout 2 from 3

Cut Handout 2 in strips, one strip for each Line ID (only one will keep the header, that is okay)

PartC: Print Handout 4 for n groups

Cut handout in half

PartD: Print Handout 5 for n groups

*NOTE: be sure to print "fit to page". Some worksheets may be too long or wide by default.*

# NOUN PROJECT ATTRIBUTES

All graphics were created by the presenters in PowerPoint or BioRender, except for the following:



Emily Akins



counloucon



Parkjisun



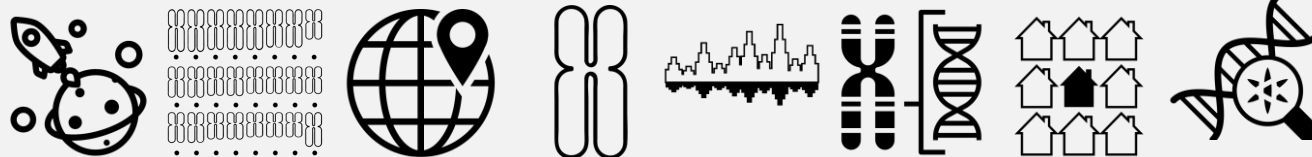
Chameleon

Wilson Joseph



Kokota

Ellie's slide icons artists: Maxim Kulikov, Rflor, Bismillah, Juan Pablo Bravo, Gregor Cresnar, Saifurrijal, Cornelia Scheitz



123rf.com

